Characterization of Hepatitis C Virus (HCV) Genotypes with Spectrum of its Viral Load
Anchal Chaurasia¹, Vinita Sangela¹, Binish Jawed¹, Mukta Sehgal¹, Vatsal Prashar³, Sachin Kumar³, Indu Bushan⁴, Guishan Kumar Dhirga⁴, Meenakshi Bahuguna¹, Narotam Sharma¹

¹Central Molecular Research Laboratory, Department of Biochemistry, Shri Guru Ram Rai Institute of Medical & Health Sciences, Patel Nagar, Dehradun-248001, Uttarakhand, India
²Kanya Gurukul Laboratory, Gurukul Kangri, University, Haridwar, Uttarakhand, India
³Gurukul Kangri Vishwavidyalaya, Haridwar, Uttarakhand, India
⁴Biotechnology Department, Maharaj Vinayak Global University, Jaipur Rajasthan, India
⁵PDM Dental College, Bahadurgarh, Haryana, India
⁶Pt. LMS (PG) College, Rishikesh, Uttarakhand, India

*Corresponding author
Dr. Narotam Sharma
Email: sharmanarotam5@gmail.com

Abstract: The Hepatitis C virus (HCV) is the most common cause of chronic liver disease and is increasing worldwide. Proposed study includes 24 clinical samples, processed for HCV RNA viral load and HCV genotyping. HCV RNA viral load ranges in between 1.00x10⁶ IU/ml to > 1.00x10⁹ IU/ml and the cases with high viral load were further processed for the genotyping. Out of 24 specimens characterized genotype 3a was prevalent in 72.72% (8/24) cases and HCV genotype 1b was in 27.27% (3/24) cases followed by HCV genotype 4a, which was in 27.27% (3/24) cases. In 13 cases target was not detected due to low viral load. 8 cases were for HCV genotype 3a of which 5 were males & 3 females, followed by HCV genotype 1b (3 males) & 4a (3 females). The genotype 3a was found in 8 (72.72%) patients. Genotype 1b was seen in 3 (27.27%) patients. Genotype 4a was seen in 3 (27.27%) patients. Genotype 2a was seen in only 1 (9.09%) patient. From the current study genotype 3a was most prevalent and can be further studied. The outcome of HCV genotyping is of almost clinical value as there are various regimens available to treat different types of HCV genotypes.

Keywords: Genotype, Sexual transmission, cirrhosis, Real Time PCR, replicating viruses, Chronic.

INTRODUCTION:
Hepatitis C virus (HCV) is an enveloped single-stranded RNA virus which appears to be distantly related to flaviviruses, although HCV is not transmitted by arthropod vectors as with other pathogens. HCV is associated with chronic liver disease and also with primary liver cancer in many countries. HCV is mostly transmitted through exposure to infective blood [3]. This may happen through transfusions of HCV-contaminated blood and blood products, contaminated injections during medical procedures, and through injection drug use [4, 6]. Sexual transmission is also possible, but is much less common. Unless successfully treated with medication, chronic Hepatitis C infection can cause other serious health problems, such as cirrhosis, liver cancer and liver failure [7, 10]. Hepatitis C is divided into six distinct genotypes throughout the world with multiple subtypes in each genotype class. In the current project the patient with HCV infection were considered & further Quantification of 5’untranslated region of the HCV genome & its genotypic characterization were determined in spectrum of its viral load [11,15].

MATERIALS AND METHODS:
A total of 24 clinical samples were considered in this study. Blood samples were taken from the infected cases of HCV and further processed at Central Molecular Research Laboratory (CMRL), Shri Guru Ram Rai Institute of Medical & Health science (SGRIM & HS), Patel Nagar, and Dehradun (U.K.) for the quantification of HCV & identification of HCV genotypes. Pre-processing of the sample collection: 5 ml EDTA Whole blood was centrifuged at 5,000 rpm for 10-15 minutes. Isolated serum was used further for RNA isolation. RNA was isolated by High Pure Viral Nucleic Acid Extraction kit. The specimens were processed for HCV viral load by Roche COBAS TaqMan 48 analyzer & the 5’untranslated region of the HCV genome was amplified for the quantification of HCV virus [16, 17]. Further HCV genotypic characterization was achieved by nested PCR targeting the current project the patient with HCV infection were considered & further Quantification of 5’untranslated region of the HCV genome & its genotypic characteristic were determined in spectrum of its viral load [11,15].
core region with genotype specific PCR primers [18, 19].

![Image of gel picture showing different genotypes of HCV at different base pairs](image.png)

Fig 1: Gel picture showing different genotypes of HCV at different base pairs

Well No.1 - HCV genotype 3a (232 bp)
Well No.2 - HCV genotype 3a (232 bp), HCV genotype 4a (99 bp)
Well No.3 - HCV genotype 1b (234 bp)
Well No.4 - HCV genotype 1b (234 bp), HCV genotype 4a (99 bp)
Well No.5 - 100 bp DNA Ladder
Well No.6 - HCV genotype 3a (232 bp)
Well No.7 - Negative (Genotype not detected due to low viral load)
Well No.8 - HCV genotype 1b (234 bp)

RESULTS:

A total of 24 clinical samples were collected from different Departments of Shri Mahant Indiresh Hospital, Patel Nagar, Dehradun U.K which includes medicine, surgery, pediatric, gastro etc were included for the proposed study. It was seen that viral load of HCV RNA ranges in between $1.00 \times 10^1$ IU/ml to $>1.00 \times 10^8$ IU/ml. Sample having the high viral load were further processed for genotyping & those less than $2.0 \times 10^5$ IU/ml were not considered because the sensitivity is low of the assay. The nested PCR yielded different amplicons for different HCV genotypes (as depicted in fig. 1). Out of 24 specimens characterized genotype 3a was prevalent in 72.72% (8/24) cases and HCV genotype 1b was in 27.27% (3/24) cases followed by HCV genotype 4a, which was in 27.27% (3/24) cases. In 13 cases target was not detected due to low viral load (limit of detection of the protocol is 25 IU/ml of HCV RNA) & all these 13 cases were not considered for further genotyping. It was also observed that in viral load ranging in between $1.00 \times 10^4$ to $1.00 \times 10^7$ IU/ml maximum number of cases were 10 and in the same spectrum HCV genotype 1b, 2a, 3a, 4a were characterized as tabulated in table no.1,2 and 3. Out of 24 samples different age groups were also considered which includes from 0-20 years, 21-40 years, 41-60 years, & above 60 years, & the number of cases in these particular age groups were 1 (Female), 8 (7 Male & 1 Female), 14 (Males), 1 (Female) respectively.

Table 1: Spectrum of HCV RNA viral load & its genotypes characterization

<table>
<thead>
<tr>
<th>S.N.o.</th>
<th>HCV RNA Titer (IU/ml)</th>
<th>No. of Cases</th>
<th>HCV Genotypes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>1a</td>
</tr>
<tr>
<td>1.</td>
<td>Target not detected</td>
<td>13</td>
<td>-</td>
</tr>
<tr>
<td>2.</td>
<td>$1.00 \times 10^3$ - $1.00 \times 10^7$</td>
<td>1</td>
<td>1b,3a,4a</td>
</tr>
<tr>
<td>3.</td>
<td>$1.00 \times 10^4$ - $1.00 \times 10^7$</td>
<td>10</td>
<td>1b,2a,3a,4a</td>
</tr>
<tr>
<td>4.</td>
<td>$&gt;1.00 \times 10^5$</td>
<td>0</td>
<td>-</td>
</tr>
<tr>
<td>Total Cases</td>
<td>= 24</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Table 2: Age wise HCV viral load and genotype distribution pattern

<table>
<thead>
<tr>
<th>Age group in years</th>
<th>No. of Cases</th>
<th>HCV genotypes</th>
</tr>
</thead>
<tbody>
<tr>
<td>0-20</td>
<td>1</td>
<td>3a</td>
</tr>
<tr>
<td>21-40</td>
<td>8</td>
<td>1b, 2a, 3a, 4a</td>
</tr>
<tr>
<td>41-60</td>
<td>14</td>
<td>1b, 3a, 4a</td>
</tr>
<tr>
<td>Above 60</td>
<td>1</td>
<td>Genotype not detected due to low viral load</td>
</tr>
</tbody>
</table>

Table 3: Percentage wise distribution of HCV genotypes

<table>
<thead>
<tr>
<th>HCV genotype</th>
<th>No. of cases</th>
<th>Percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td>1a</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>1b</td>
<td>3</td>
<td>27.27%</td>
</tr>
<tr>
<td>2a</td>
<td>1</td>
<td>9.09%</td>
</tr>
<tr>
<td>2b</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>3a</td>
<td>8</td>
<td>72.72%</td>
</tr>
<tr>
<td>3b</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>4a</td>
<td>3</td>
<td>27.27%</td>
</tr>
</tbody>
</table>

Fig 2: Amplification plot for HCV RNA viral load (Case= High HCV viral titer).

Fig 3: Amplification plot for HCV RNA viral load (Case= Low HCV viral titer)
DISCUSSION:
Differences among HCV genotypes in geographic distributions have provided investigators with an epidemiological marker that can be used to find the source of HCV infection in a given population & for further prognosis [20, 21]. The current study includes the collection of 24 specimens which were confirmed positive for HCV by serological findings. As the hospital includes the involvement of patients from hilly areas of Uttarakhand & Himachal Pradesh along with some parts of West U.P. Out of 11 samples which was processed for genotyping, 8 cases were for HCV genotype 3a of which 5 were males & 3 females, followed by HCV genotype 1b (3 males) & 4a (3 males). The genotype 3a was observed in 8 (72.72%) patients. Genotype 1b was seen in 3 (27.27%) patients. Genotype 4a was seen in 3 (27.27%) patients. Genotype 2a was seen in only 1 (9.09%) patient. Our findings include the prevalence of HCV genotype 3a in the clinical isolates, although HCV genotypes 1, 2 and 3 were also detected. From the current study genotype 3a was most prevalent and was present in males (the reason is not clear but it may be due to injection, drug abuse, needle stick accidents and transfusion of blood, intravenous drug users & tattooing etc). Similar study was also done by Pushpa Latha Manjunatha et al.; in 2015 and Chakravarti, Anita, Gaurav Dogra et al.; in 2011 [22-25]. This work can be further evaluated to study the exact HCV genotyping characterization in the Northern Indian population with more number of samples because this study was limited to the 24 samples only.

CONCLUSION:
The outcome of HCV genotyping is of almost clinical value as there are various regimens available to treat different types of HCV genotypes like Simeprevir, Sofosbuvir etc for genotype 1. Sofosbuvir/R for genotype 2 & Sofosbuvir/R for geno type 3. There are various alternative therapies also available to treat Hepatitis C infection like Milk Thistle, Green tea extract, Glycyrrhizin but currently there is no vaccine available to prevent the Hepatitis C infection. The distribution of HCV genotypes vary according to the geographical region (26–28). The HCV genotype 3a is one of the most replicating viruses known to damage hepatic cells & thus requires proper line of treatment thoroughly during the diagnosis. Further the study can be done with more number of clinical isolates to get the exact epidemiological profiling of HCV genotypes.

Conflict of Interest: None

Acknowledgment: The authors are grateful to Honourable Chairman, Shri Guru Ram Rai Education Mission for his kind support and guidance.

REFERENCES:
4. Berry KE, Waghray S, Mortimer SA, Bai Y, Doudna JA. Crystal structure of the HCV IRES central domain reveals strategy for start-codon