INTRODUCTION

Plants have always been a part of medicinal science from the beginning of human civilization to the present modern world of synthetic medicines. Even in the presence of variety of effective synthetic drugs, use of medicinal plants for maintaining human health has acquired a lot of importance in the present era [1]. There is a global interest in non-synthetic, natural drugs derived from plant sources, because of low cost, non-toxic nature and availability. Many plants with antioxidant potential possess flavonoids and phenolic compounds [2]. Free radical reactions have been implicated in the pathology of many human disease including atherosclerosis, ischemic heart disease, the aging process, inflammation diabetes and other conditions [3].

The research for medicinal plants discovered various common plants having distinguishable medicinal properties, among which one is Bougainvillea glabra. Bougainvillea glabra is an ornamental flowering plant from the genus of bougainvillea; family Nyctaginaceae [4] and a native to Brazil. The genus bougainvillea has eighteen species, of which B. spectabilis, B. glabra and B. Peruvian [5] are horticultural important. Bougainvillea glabra is a woody cliver with thorny thin stems and long branches; also it has papery bracts and smooth leaves, which grows to more than 10 meters of height [6].

B. glabra is one that is great for container plantings and produce brilliant colors. The plant is perfect for arbors and draped long fences where it creates a security barrier by its thorns and twiggy growth [7]. Body has itself antioxidant system, which reacts with reactive species and neutralizes them. This natural antioxidant system includes enzymes like catalase, superoxide dismutase and glutathione, which protect the body from free radical species and prevent oxidative stress [8]. Synthetic antioxidant like butylated hydroxyl toluene and butylated hydroxyl anisole are carcinogenic in nature. So, there arises a need for natural antioxidant [9]. The antioxidant activity of the hydroalcoholic, acetone extracts of the leaves were evaluated for this plant [10]. With this background, the aim of the present study was to determine the possible phytochemical and antioxidant activity of chloroform extract of Bougainvillea glabra choice leaves.

MATERIALS AND METHODS

Plant Material

Plants were collected form Thanjavur District of Tamilnadu. The botanical identity of the plant of Bougainvillea glabra was confirmed by Dr. John Brito, Rapinet Herbarium. St.Joseph’s College, Thiruchirappalli.

Preparation of Extract

The powder (5kg) was extracted with 95% ethanol at room temperature for seven days. The extracts were filtered and concentrated under reduced pressure in a rotary evaporator and for further fractionated successively with n-hexane, chloroform, ethyl acetate, acetone, ethanol, butanol and methanol solvents were then removed under reduced pressure. From the fractionation chloroform extract were subjected to screening of phytochemical and antioxidant activity.
Phyto Chemical Analysis
Phytochemical analysis involves the qualitative analysis of herbal plants. The preliminary qualitative tests have been attempted in Bougainvillea glabra to find out the presence or absence of certain bioactive Compounds. Chemical tests were carried out on the chloroform extract using standard procedures to identify the constituents as described by Harborne [11].

Test for Tannins
About 0.5g of the dried powdered samples was boiled in 20ml of water in a test tube and then filtered. A few drops of 0.1% ferric chloride were added. Presence of brownish color indicated the presence of tannins.

Test for Phlobatannins
Extract boiled with 1% aqueous hydrochloric acid. No red precipitate is formed indicated absence of phlobatannins.

Test for Saponin
About 2g of the powdered sample was boiled in 20ml of distilled water in a water bath and filtered. 10ml of the filtrate was mixed with 5ml of distilled water and shaken vigorously for a stable persistent froth. No change in solution. Saponin is absent.

Test for Flavonoids
Few drops of 1% aluminum solution were added to a portion of extract. A yellow colouration was observed indicating the presence of flavonoids.

Test for Oil and Fat
Stain test
Small Quantity of extract was pressed between two filter papers. An oily stain on filter paper indicates the presence of fixed oil.

Test for Steroids
2ml of acetic anhydride was added to 0.5g ethanolic extract with 2ml sulphuric acid. The color changed from violet to blue indicating the presence of steroids.

Test for Terpenoids (Salkowski Test)
5ml of extract was mixed in 2ml of chloroform, and 3ml of conc. Sulphuric acid was carefully added to form a layer. A reddish brown colouration of the interface was formed to show positive results for the presence of terpenoids.

Test for Alkaloid
Mayer’s Test
0.5 ml of extract is added with 1 ml of mayer reagent. Absence of yellow colour indicated absence of alkaloid.

Dragandroff’s test
0.5ml of extract is added with 1ml of dragandroff’s reagent. Absence of orange red colour indicated absence of alkaloid.

Wagner’s Test
0.5ml Sample is treated with 1 ml of wagner reagent. No change in colour indicated absence of alkaloids.

Antioxidant Activity
DPPH method
0.1 ml of the chloroform extract was taken in test tubes. 6 ml of DPPH (diphenyl picryl hydrazyl) solution was added and the tubes kept in dark for one hour. The color was read at 517 nm. The difference in the Optical density of DPPH solution and DPPH solution + sample was calculated. The decrease in OD with sample addition is used for calculation of the antioxidant activity. The activity was compared with BHT (b tylated hydroxytoluene) standard [12]. Free radical scavenging activity was expressed as the inhibition percentage calculated using the formula.

\[
\text{Percentage of antioxidant activity} = \frac{[\text{Absorbance of control} - \text{Absorbance of sample}]}{\text{Absorbance of sample}} \times 100
\]

Reducing power assay
Reducing power assay of sample was done using published protocols [13]. 1 ml of chloroform extract was mixed with phosphate buffer (2.4 ml 0.2 M pH 6.6) and potassium ferricyanide (2.5 ml). The mixture was incubated at 50°C for 20 minutes. A portion (2.5 ml) of trichloroacetic acid (10%) was added to the mixture, which was then centrifuged at 3000 rpm for 10 minutes. The upper layer of solution (2.5 ml) was mixed with distilled water (2.5 ml) and Ferric chloride (0.5 ml, 0.1%) and absorbance measured at 700 nm. Increased absorbance of the reaction mixture indicates stronger reducing power. The activity was compared with BHT standard. Scavenging activity was expressed as the inhibition percentage calculated using the formula.

\[
\text{Percentage of antioxidant activity} = \frac{[\text{Absorbance of control} - \text{Absorbance of sample}]}{\text{Absorbance of sample}} \times 100
\]

Hydrogen peroxide scavenging activity
To determine the Hydrogen peroxide assay of chloroform extract by Umamaheswari and Chatterjee Method [14]. Hydrogen peroxide solution (2 mM/L) was prepared with standard phosphate buffer (pH 7.4). Different concentration of the extracts in distilled water was added to 0.6 ml of hydrogen peroxide solution. Absorbance was determined at 230 nm after 10 minutes against a blank solution containing phosphate buffer without hydrogen peroxide. The inhibition was calculated. BHT was used as standard. Scavenging activity was expressed as the inhibition percentage calculated using the formula.
Percentage of antioxidant activity = \[ \frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of sample}} \times 100 \]

RESULTS AND DISCUSSION

Table 1: Qualitative analysis of phytochemicals of *Bougainvillea glabra* choicy leaves.

<table>
<thead>
<tr>
<th>Sl. No.</th>
<th>Phytochemical</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Alkaloids</td>
<td>Absent</td>
</tr>
<tr>
<td>2</td>
<td>Pholobtannin</td>
<td>Absent</td>
</tr>
<tr>
<td>3</td>
<td>Tannin</td>
<td>Present</td>
</tr>
<tr>
<td>4</td>
<td>Terpenoid</td>
<td>Present</td>
</tr>
<tr>
<td>5</td>
<td>Saponin</td>
<td>Absent</td>
</tr>
<tr>
<td>6</td>
<td>Flavonoids</td>
<td>Present</td>
</tr>
<tr>
<td>7</td>
<td>Steroids</td>
<td>Present</td>
</tr>
<tr>
<td>8</td>
<td>Fatty acid</td>
<td>present</td>
</tr>
</tbody>
</table>

Table 2: Antioxidant activity of chloroform extract of *Bougainvillea glabra* choicy leaves.

<table>
<thead>
<tr>
<th>Sl no.</th>
<th>Method</th>
<th>Control Absorbance</th>
<th>Sample Absorbance</th>
<th>% Antioxidant activity</th>
<th>Standard (BHT) (0.01 mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>DPPH assay</td>
<td>1.586</td>
<td>0.421</td>
<td>73.45</td>
<td>78.74</td>
</tr>
<tr>
<td>2</td>
<td>Reducing power assay</td>
<td>1.007</td>
<td>0.538</td>
<td>46.57</td>
<td>69.72</td>
</tr>
<tr>
<td>3</td>
<td>Hydrogen Peroxide</td>
<td>1.389</td>
<td>0.983</td>
<td>29.22</td>
<td>45.92</td>
</tr>
</tbody>
</table>

The antioxidant activity showed the inhibition percentage is 73.45 %, 46.57 % and 29.22 % respectively. The results observed from chloroform extract of bougainvillea glabra choicy leaves shows higher antioxidant potential and compared to three methods DPPH assay observed high antioxidant activity. The results were compared to the standard BHT, it was only slight difference has been noted. The WHO estimated that 80% of the population of developing countries still relies on traditional medicine, mostly plant drugs for their primary health care needs. Hence, there is an urgent need to study the screening of antioxidant properties of herbs which will be helpful in the treatment of several diseases [15].

Antioxidants are an inhibitor of the process of oxidation, even at relatively small concentration and thus have diverse physiological role in the body. Antioxidants may be synthetic or natural. Synthetic antioxidants such as BHT and BHA have recently been reported to be dangerous for human health. Thus, the search for effective, non-toxic natural compound with antioxidative activity has been intensified in recent years [16]. On the basis of our results, bougainvillea appears to have potential for treatment of oxidative stress related diseases. It should, however, be explored as a functional medicinal plant for isolating the active ingredients along with animal studies in vivo.

Acknowledgement
I wish to express my deep sense of gratitude and most sincere thanks to S. Kumaravel, senior scientist, Indian Institute of Crop Processing Technology, Thanjavur for providing all the facilities and support to finish my work.

REFERENCES
8. Elumalai A, Chinna Eswariah M, Vidhyulatha Chowdary CH, Ranjit Kumar, Anusha M and...


